Protocol for Meconium Homogenization in the Bullet Blender®

The protocol described in this document is for the use of the Bullet Blender[®] for the homogenization of human meconium. This protocol does not specify a particular buffer - you may choose which is most appropriate for your downstream application (nucleic acid isolation, protein extraction, etc.).

Materials

meconium, Bullet Blender[®], homogenization buffer, pipettor, microcentrifuge tubes and Red bead lysis kit/Pink bead lysis kit/0.5mm zirconium oxide beads (part number ZrOB05).

Instructions

- 1. Weigh a mass of meconium (10mg-300mg) to be homogenized. Try to keep the walls of the tube as clean as possible, and ensure the meconium sits as low in the tube as possible.
- 2. a. Samples 100mg or greater
 Place the sample in Red bead lysis kit tube.
 - b. Samples less than 100mg
 Place the sample in Pink bead lysis kit tube.
 - c. Alternate protocol step for bulk beads

 Place sample in microcentrifuge tube and add beads to the tube. Use a volume of beads equal to the mass of tissue. **NOTE:** $100 \text{mg} \cong 100 \mu\text{L}$.
- 3. Securely close the microcentrifuge tubes.
- 4. Place tubes into the Bullet Blender®.
- 5. Set controls for **SPEED 8** and **TIME 5** minutes. Press **Start**.
- 6. After the run, remove tubes from the instrument.
- 7. Verify that all samples have a uniform appearance. A smooth semi-liquid should exist. If homogenization is unsatisfactory, run for another five minutes at speed 8.
- 8. Proceed with your downstream application.

SAFETY NOTE!!!

When using a centrifuge to separate your homogenate from the debris and beads, make sure your tubes are balanced.

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