# Homogenization in the Bullet Blender<sup>®</sup> 50 Protocol for Hepatic (Liver) Tissue

The protocol described in this document is for the use of the Bullet Blender<sup>®</sup> 50 for the homogenization of Liver/Hepatic Tissue. This protocol does not specify a particular buffer - you may choose which is most appropriate for your downstream application (nucleic acid isolation, protein extraction, etc.).

#### Materials Required:

liver tissue , Bullet Blender<sup>®</sup> 50, homogenization buffer, pipettor, 50mL skirted centrifuge tubes (Axygen<sup>®</sup> or Corning<sup>®</sup> brand), 4.8mm stainless steel beads (part number SSB48) or 3.2mm stainless steel beads (part number SSB32).

#### Instructions

- 1. Cut liver tissue into appropriately sized pieces for analysis (0.1g 3.5g) and place into a 50mL centrifuge tube.
- **2. OPTIONAL:** If desired, wash the tissue 3x with 5mL PBS to remove blood and other contaminants from the tissue.
- **3.** Add a mass of stainless steel beads to the tube equal to approximately 6x the mass of your sample.
- **4.** Add 0.2 mL to 7mL buffer (2 volumes of buffer for every mass of sample).
- **5.** Screw caps onto centrifuge tubes **TIGHTLY**.
- **6.** Place tubes into the Bullet Blender<sup>®</sup> 50.
- 7. Set controls for **SPEED 8** and **TIME 12** minutes.
- **8.** Remove tubes from the instrument.
- **9.** Visually inspect samples, if homogenization is unsatisfactory, run for another six minutes at the **SPEED 9**.
- **10.** Proceed with your downstream application.

## SAFETY NOTE!!!

### When using a centrifuge to separate your homogenate from the debris and beads, make sure your tubes are balanced.